#### **RESEARCH ARTICLE**

# The use of spray-drying to enhance celecoxib solubility

Ehab A. Fouad<sup>1,2</sup>, Mahmoud EL-Badry<sup>1,2</sup>, Gamal M. Mahrous<sup>1</sup>, Fars K. Alanazi<sup>1,3</sup>, Steven H. Neau<sup>4</sup>, and Ibrahim A. Alsarra<sup>1,5</sup>

<sup>1</sup>Department of Pharmaceutics, College of Pharmacy, King Saud University, Riyadh, Saudi Arabia, <sup>2</sup>Department of Pharmaceutics, Faculty of Pharmacy, Assiut University, Assiut, Egypt, <sup>3</sup>Kayyali Chair for Pharmaceutical Industry, College of Pharmacy, King Saud University, Riyadh, Saudi Arabia, <sup>4</sup>Department of Pharmaceutical Sciences, Philadelphia College of Pharmacy, University of the Sciences, Philadelphia, PA, USA, and 5Center of Excellence in Biotechnology Research, King Saud University, Riyadh, Saudi Arabia

#### Abstract

The present research investigates the enhancement of the dissolution rate of celecoxib by using spray-drying to prepare a solid dispersion with various polymers, namely Kollicoat IR° (Kollicoat), polyvinyl alcohol (PVA) 22000, or polyethylene glycol 6000 (PEG). The investigated drug-to-polymer mass ratios were 1:1, 1:2, and 1:4 by weight. Hydroalcoholic or methylene chloride solvent systems were used. The obtained yields ranged from 65% to 78%, whereas the entrapment efficiencies were between 68% and 82%. The results revealed an increase in the dissolution rate of the prepared particles up to 200% within 20 min. The prepared particles were investigated using differential scanning calorimetry, scanning electron microscopy, X-ray diffraction, and Fourier transform infrared spectroscopy. The increased dissolution rate was attributed to hydrogen bond formation between celecoxib and each polymer together with the reduced size of the formed particles offering a greater overall surface area. It was concluded that spray-drying may be considered a successful one-step technique to improve the dissolution rate of celecoxib when using Kollicoat, PVA, or PEG as the carrier polymer.

Keywords: Spray-drying, celecoxib, Kollicoat IR®, polyvinyl alcohol, polyethylene glycol, dissolution rate

### Introduction

Oral drug administration is perhaps the most appealing route for drug delivery1. The poor solubility of drugs is a common concern since, to achieve good oral bioavailability, drugs must first dissolve to allow their absorption. Celecoxib, 4-[5-(4-methylphenyl)-3-(trifluoromethyl)-1H-pyrazol-1-yl]benzenesulfonamide, is a selective cyclooxygenase (COX-2) inhibitor that is widely prescribed for pain and inflammation<sup>2</sup>. It inhibits the conversion of arachidonic acid to the prostaglandins that mediate pain and inflammation while having no effect on the formation of the prostaglandins that mediate normal homeostasis in the gastrointestinal (GI) tract<sup>3</sup>. It is also used in the treatment of arthropathies and adenomatous polyps<sup>4</sup> and in dentistry<sup>5</sup>. It has a comparable efficacy and superior gastric tolerability6 and is safer when compared with conventional nonsteroidal anti-inflammatory drugs<sup>7</sup>. Celecoxib has a p $K_a$  of 11.18, with a low aqueous solubility of 3–7  $\mu$ g/mL<sup>9</sup>. With a p $K_a$  of 11.1, celecoxib is unionized at physiological pH. It is not surprising that celecoxib is well-absorbed from the GI tract considering its logP of 2.82 between buffer and octanol at pH 7.4 and room temperature8,10.

After oral solution dosing in dogs, celecoxib was rapidly absorbed and reached maximum concentrations by 1h; absorption was delayed 1 to 2h when administered as a solid. The absolute bioavailability of celecoxib was higher when given as a solution (64-88%) compared with a capsule dosage form (22-40%)9. According to the biopharmaceutical classification system, celecoxib can be categorized as class II that includes poorly water-soluble drugs with high GI permeability<sup>11</sup>.

In addition, celecoxib has handling problems due to its needle-shaped crystals12 that have high surface

Address for Correspondence: Prof. Ibrahim A. Alsarra, Department of Pharmaceutics, College of Pharmacy, King Saud University, P.O. Box 2457, Riyadh 11451, Saudi Arabia. Tel: +(966)-1-4677504. Fax: +(966)-1-4676363. E-mail: ialsarra@ksu.edu.sa



energy. Modification of the crystal habit or solid structure of celecoxib would be advantageous. Celecoxib can form a metastable polymorph, but the difference in melting point from that of the stable polymorph is negligible and it is capable of reverting to the stable form12.

Several formulation approaches have been attempted to improve the dissolution properties of celecoxib for faster onset of action and improved bioavailability. Examples include formulation of self-microemulsifying drug delivery systems (SMEDDS)13, solid dispersions (SD)<sup>14</sup>, complexation with β-cyclodextrins<sup>15</sup>, and manipulation of the solid state of the drug16,17. However, some of these formulations employ or contain a high concentration of surfactants (20-60%), which may eventually compromise their uses in chronic treatment<sup>18</sup>.

Kollicoat IR®, a polyvinyl alcohol (PVA)-polyethylene glycol (PEG) graft copolymer, is a pharmaceutical excipient that was developed as a coating polymer for immediate release tablets19. PVA has good film-forming properties and the PEG portion acts as an internal plasticizer. The molecule is hydrophilic and readily soluble in water. As its structure is non-ionic, its solubility does not change when pH is altered along the GI tract.

The major drawback of celecoxib is its poor aqueous solubility and consequently its slow dissolution in gastric fluid. Hence, we sought to enhance these properties by using spray-drying to prepare binary mixtures with Kollicoat IR® (Kollicoat), PVA 22000, and PEG 6000.

# Materials and methods

Celecoxib was kindly donated by Searle (Augusta, GA). Kollicoat IR® (Molecular weight = 45,000 Da) was obtained from BASF (Ludwigshafen, Germany). PVA with a molecular weight of 22,000 and PEG 6000 were purchased from BDH Chemicals Ltd. (Poole, England). Silicon dioxide was obtained from Serva Fine Biochemical GmbH Co. (Heidelberg, Germany). Sodium lauryl sulfate was obtained from E-Merck (Darmstadt, Germany). All other chemicals were of reagent grade.

# Preparation of binary mixtures

Physical mixtures (PMs) were prepared by mixing celecoxib and each of the carriers in a mortar. The ratio of celecoxib to the carrier used was 1:1 or 1:4 by weight.

# Spray-dried binary systems using Kollicoat IR® or PVA

Appropriate mass ratios (1:1, 1:2, and 1:4) of celecoxib and either Kollicoat IR® or PVA were prepared in a 2:1 v/v hydroalcoholic solution where celecoxib was completely dissolved in ethanol and Kollicoat or PVA was dissolved in distilled water. The aqueous solution was added gradually to the ethanolic drug solution with subsequent vigorous stirring for 1 h to assure equilibrium. The resultant suspension was spray-dried in a Mini Spray-Dryer B-290 (Büchi Labortechnik AG, Flawil, Switzerland) with the following conditions: inlet temperature 130°C, outlet temperature 60-65°C, suspension flow rate 5 mL/ min, air flow rate 40-50 m<sup>3</sup>/h, and atomizing air pressure 1.0-1.1 bar. The batch size of the prepared ratios was 10 g each.

# Spray-dried binary systems using PEG

Celecoxib in combination with PEG 6000 in different mass ratios (1:1, 1:2, and 1:4) was dissolved in 100 mL of dichloromethane. To these clear solutions, silicon dioxide (2% w/v) was slowly added to obtain uniform suspensions. The suspension was spray-dried in the Büchi mini spray-dryer with the following conditions: inlet temperature 50°C, outlet temperature 30°C, solution flow rate 5 mL/min, air flow rate 40-50 m<sup>3</sup>/h, and atomizing air pressure 1.0-1.1 bar. The batch size was again 10 g.

# Microparticle characterization

# Drug loading

The efficiency with which celecoxib was entrapped in the particle powder after spray-drying was determined as the mass ratio of the entrapped drug to the theoretical amount of celecoxib used in the preparation. Spraydried particles equivalent to 5 mg of drug were accurately weighed and dissolved in a suitable quantity of ethanol. The drug content was determined spectrophotometrically at 252 nm18.

#### Morphological analysis

The morphological characteristics of spray-dried particles were observed by scanning electron microscopy. The samples were sputter-coated with a thin gold palladium layer under an argon atmosphere using a sputter module in a high-vacuum evaporator. The coated samples were then scanned and photomicrographs were taken with a JSM-1600 scanning electron microscope (Jeol, Tokyo, Japan).

#### Differential scanning calorimetry

Calorimetric studies of the drug and the prepared microparticles were performed using a DSC-60 (Shimadzu, Kyoto, Japan). The 3-5 mg samples were placed in hermetically sealed aluminum pans. A 10°C/ min scanning rate was used over the 25-200°C temperature range. Indium was used as the temperature and enthalpy standard.

### Powder X-ray diffractometry

Powder X-ray diffraction patterns of the drug, polymers, and the prepared spray-dried microparticles were generated using a wide-angle Rigaku Ultima IV X-ray diffractometer (Rigaku Corporation, Tokyo, Japan). The instrument was operated on the  $2\theta$  scale. The angular range was 10° to 50° (20) and counts were accumulated for 1 sec at each step.

#### Fourier transform infrared spectroscopy

Infrared spectra were recorded on a PerkinElmer spectrum BX FTIR (PerkinElmer, Waltham, MA). Samples were prepared as KBr pellets and scanned against a blank KBr pellet at wave numbers ranging from 4000 to 650 cm<sup>-1</sup> with resolution of 1.0 cm<sup>-1</sup>.

# Dissolution study for microparticles

The drug dissolution studies were performed using a Caleva Model 85T (Philips, Maidstone, UK) USP dissolution apparatus II at 50 rpm. A continuous automated monitoring system, consisting of an IBM PK 8620 computer and PU 8605/60 dissolution test software, was used with a Philips Model PU 8620 Vis/UV/NIR single beam eight cell spectrophotometer and a Watson-Marlow peristaltic pump. Each vessel, containing 900 mL of 1.0% sodium lauryl sulfate20 in demineralized water, was maintained at 37.0 ± 0.5 °C. Microparticles containing 100 mg of drug were sieved through a 200-µm sieve and then dispersed in the dissolution medium. Dissolved drug was determined spectrophotometrically at 252 nm. The dissolution experiments were conducted in triplicate and the means of the percent of drug dissolved were calculated.

### **Results and discussion**

Carriers for the production of SDs via spray-drying should be chemically and physically stable with a low melting point and solubility in the solvent systems whether aqueous or nonaqueous. Further requisites are chemical compatibility with the drug, miscibility with the drug in the liquid state, and the ability to increase the solubility of the drug in an aqueous medium. These properties can be achieved with water-soluble excipients that exhibit rapid dissolution themselves. Excipients with a high number of functional groups are typically able to engage in intermolecular hydrogen bonds, van der Waals forces, and ionic interactions.

#### Differential scanning calorimetry

Differential scanning calorimetry (DSC) thermograms for crystalline celecoxib, the 1:4 PM, and SD at mass ratios of 1:1, 1:2, and 1:4 celecoxib:polymer are shown in Figures 1-3. A loss of drug crystallinity increases its dissolution rate. The pure drug melted at 159.49°C (Figure 1), which agrees well with the literature value of 160.8°C21 and the PM with PVA had essentially the same onset temperature. The endothermic peak for spray-dried particles shifted to 157.8°C, 157.8°C, and 155.3°C for the mass ratios 1:1, 1:2, and 1:4, respectively, indicating drug-polymer interactions. The enthalpy likewise decreased compared with that of pure celecoxib, yielding 6.77, 7.64, 4.95, and 3.4 J/g for the PM and spray-dried particles at mass ratios of 1:1, 1:2, and 1:4, respectively. These are dramatic reductions in comparison with the 82.8 J/g enthalpy of fusion of celecoxib<sup>18,21,22</sup>. With PEG 6000, the endothermic peak for celecoxib disappeared in the thermogram for each combination, whether the drug was in a PM or spraydried particles (Figure 2). The specific PEG endothermic peak decreased in intensity with the spray-dried particles. The thermograms for celecoxib and Kollicoat mixtures showed celecoxib endothermic peaks at

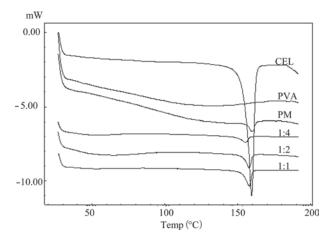


Figure 1. Differential scanning calorimetric (DSC) thermograms of celecoxib, polyvinyl alcohol (PVA) 22000, their physical mixture, and various spray-dried particles.

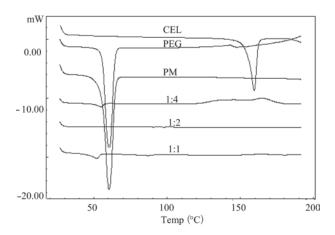


Figure 2. Differential scanning calorimetric (DSC) thermograms of celecoxib, polyethylene glycol (PEG) 6000, their physical mixture, and various spray-dried particles.

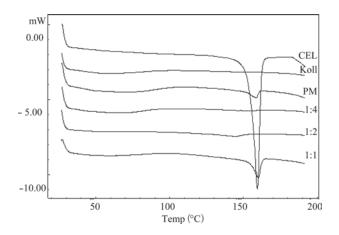


Figure 3. Differential scanning calorimetric (DSC) thermograms of celecoxib, Kollicoat, their physical mixture, and various spraydried particles.



158.6°C, 160.1°C, 144.6°C, and 156.1°C for the PM, and spray-dried particles at mass ratios of 1:1, 1:2, and 1:4, respectively (Figure 3). The peaks showed decreasing enthalpy of 17.11, 12.5, 8.62, and 0.23 J/g for the PM, and spray-dried particles at mass ratios of 1:1, 1:2, and 1:4, respectively.

Each of the polymers either softens when heated above its glass transition temperature or melts at a temperature below that of celecoxib and this may lead to dissolution of the drug in the polymer. Accordingly, this will lead to disappearance of the drug melting endotherm in the DSC thermogram of the PM.

Using calorimetric data, the solubility factor for celecoxib in the different polymers was calculated (Table 1) by dividing the enthalpy obtained from the endothermic peak for celecoxib in prepared particles by that for pure celecoxib. As the solubility factor decreases, the solubility of celecoxib in the polymer increases. If it reaches one, celecoxib did not dissolve in the polymer. The data show that the drug is soluble in PEG at each of the tested mass ratios. Particles containing PVA and Kollicoat showed increases in drug solubility as the polymer level

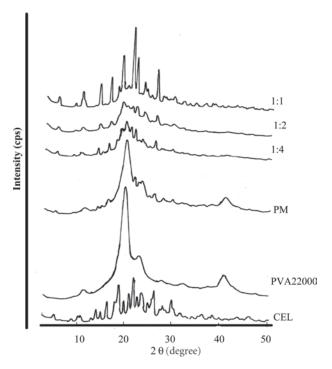


Figure 4. X-ray diffractograms of the spray-dried dispersions of 1:1, 1:2, and 1:4 mass ratios with polyvinyl alcohol (PVA), their physical mixture, PVA 22000 alone, and celecoxib alone.

Table 1. Solubility factor of celecoxib in different polymers.

	Polymers		
	Polyvinyl alcohol	Polyethylene glycol (PEG)	
Combination	(PVA) 22000	6000	Kollicoat IR®
Physical mixture	0.29	0	1.00
SD 1:1	0.39	0	0.42
SD 1:2	0.18	0	0.20
SD 1:4	0.12	0	0.01

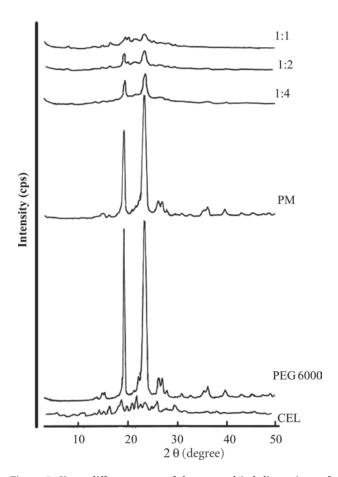


Figure 5. X-ray diffractograms of the spray-dried dispersions of 1:1, 1:2, and 1:4 mass ratios with polyethylene glycol (PEG), their physical mixture, PEG 6000 alone, and celecoxib alone.

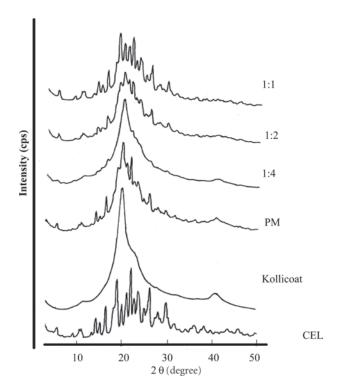


Figure 6. X-ray diffractograms of the spray-dried dispersions of 1:1, 1:2, and 1:4 mass ratios with Kollicoat, their physical mixture, Kollicoat alone, and celecoxib alone.

in the particle increased. Celecoxib dissolved to a limited extent in the PM with PVA, dissolved completely with PEG, and proved to be insoluble with Kollicoat in the PM. The spray-drying process resulted in an increase in the solubility factor in going from the PM to the SD of celecoxib with PVA or Kollicoat. This may be attributed to a polymer effect on the celecoxib crystal form or an interaction of the drug with the polymer. As the drug level increases in the mass ratio, the solubility factor decreases because surplus crystalline drug is present.

# **Powder X-ray diffraction**

Powder X-ray diffractograms of crystalline celecoxib, the 1:4 PM, and the SD at 1:1, 1:2, 1:4 mass ratios are presented in Figures 4-6. Characteristic peaks of celecoxib appear at 5.6°, 10.6°, and 16.4° 20 corresponding to reported crystal lattice parameters for celecoxib at 5.0°, 10.1°, and 16.8°  $2\theta^{23}$ . For PVA, diffraction peaks appear at 19.7° and 40.7° 2θ (Figure 4) corresponding to reported peaks at 19.4° and 40.4°  $2\theta^{24}$ . The diffraction pattern for PEG exhibited the two strong characteristic peaks at 19.3° and 23.5° 2θ (Figure 5) that have been reported<sup>25</sup>. For Kollicoat, the diffraction pattern showed a strong broad peak at 19.6 with a shoulder at 23.5° 2θ representing the overlapping peaks for PVA and PEG chains present in Kollicoat, and a small one at 40.7° 2θ corresponding to the peak for the PVA portion (Figure 6).

With the exception of PEG, the PM diffraction patterns still reveal peaks characteristic of crystalline celecoxib,

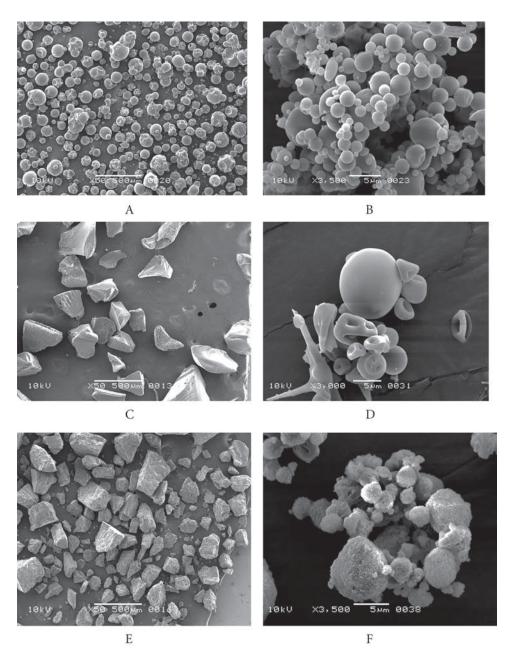


Figure 7. SEM photomicrographs of Kollicoat IR (A), polyvinyl alcohol (PVA) 22000 (C), polyethylene glycol (PEG) 6000 (E), and spraydried particles in 1:4 ratio (B, D, F), respectively.



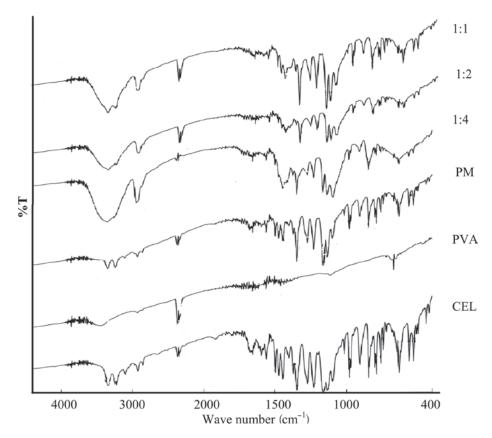


Figure 8. FTIR spectra of the various spray-dried dispersions with mass ratios of 1:1, 1:2, and 1:4 with polyvinyl alcohol (PVA), the physical mixture, PVA 22000 alone, and celecoxib alone.

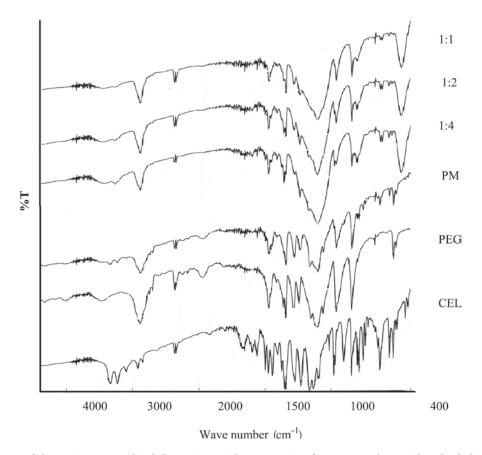


Figure 9. FTIR spectra of the various spray-dried dispersions with mass ratios of 1:1, 1:2, and 1:4 with polyethylene glycol (PEG), the physical mixture, PEG 6000 alone, and celecoxib alone.

although the intensity of the peaks is reduced. The PEG peaks in the 1:4 mass ratio PM or the 1:4 mass ratio SD are intense and the drug peaks are likely of negligible intensity in comparison with those of PEG or are undetectable, respectively (Figure 5). For each SD with a 1:1 mass ratio, the polymer and celecoxib peaks both exist and diminish in intensity, revealing that at least part of the celecoxib present is crystalline in the SD at this mass ratio. Drug peaks in the diffractograms continue to diminish as the polymer level in the mass ratio is increased.

## Scanning electron microscopy

The scanning electron micrographs for PM and SD particles with PVA, PEG, and Kollicoat are presented in Figure 7. Spray-drying solutions of celecoxib with PVA formed rings and spherical particles both with a smooth surface (Figure 7d), whereas with Kollicoat smaller spherical particles with a smooth surface were produced (Figure 7b). For SDs with PEG, the particles appear larger with a nearly spherical shape and a rough surface likely due to the presence of silicon dioxide (Figure 7f).

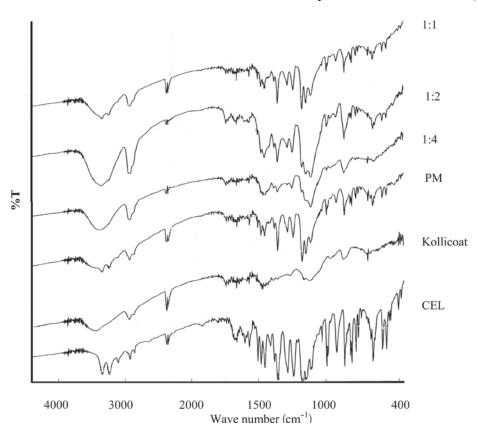


Figure 10. FTIR spectra of the various spray-dried dispersions with mass ratios of 1:1, 1:2, and 1:4 with Kollicoat, the physical mixture, Kollicoat alone, and celecoxib alone.

Group	Celecoxib	Physical mixture	SD 1:1	SD 1:2	SD 1:4	
Polyvinyl alcohol (	PVA) 22000					
$\mathrm{NH}_{\scriptscriptstyle 2}\mathrm{str}$	3340.5	3340.7	3342	3346	3345.9	
$\mathrm{NH}_{_2}\mathrm{str}$	3234.5	3234.1	_	_	_	
S=O sym str	1348.7	1348.3	1348.2	1348.9	1348.2	
S=O asym str	1165.5	1165.2	1164.9	1165.7	1165.4	
Polyethylene glycol (PEG) 6000						
$\mathrm{NH}_{_2}\mathrm{str}$	3340.5	3341.5		_	_	
$\mathrm{NH}_{\scriptscriptstyle 2}\mathrm{str}$	3234.5	_	3266.1	_	_	
S=O sym str	1348.7	1348.3	1343.2	1343.3	1343.2	
S=O asym str	1165.5	1165.5	_	_	_	
Kollicoat IR®						
$\mathrm{NH}_{_2}\mathrm{str}$	3340.5	3342.0	3342.7	3341.6	3379.8	
$\mathrm{NH}_{_2}\mathrm{str}$	3234.5	_	_	_	_	
S=O sym str	1348.7	1343.3	1348.1	1348.1	1348.3	
S=O asym str	1165.5	1164.9	1165.6	1165.4	1165.2	



#### FTIR spectroscopy

Celecoxib has certain characteristic peaks that are evident in Figures 8-10. These include bands at 3340.5 and 3234.5 cm<sup>-1</sup> that correspond to N-H stretching in the SO<sub>2</sub>NH<sub>2</sub> group, and 1348.7 and 1165.5 cm<sup>-1</sup> for the S=O asymmetric and symmetric stretching<sup>21</sup>. The manner in which these bands change when drug is physically mixed with the polymers or existing in the different solid dispersions with the polymers is presented in Table 2. The PEG bands corresponding to the CH<sub>2</sub> was at 1370 cm<sup>-1</sup> and the C-O-C asymmetric stretching at about 1100 cm<sup>-1</sup> overshadow or obscure the 1348.7 and 1165.5 cm<sup>-1</sup> band for celecoxib and make interpretation difficult. However, in the 1:1 mass ratio SD where the influence of drug on PEG can be greatest, it is apparent that the PEG O-H stretching vibration at about 3400 cm<sup>-1</sup> diminishes in intensity and the two celecoxib bands at 3340.5 and 3234.5 also diminish and broaden, suggesting that N-H from the drug engages in an interaction with the O-H of PEG, likely by hydrogen bonding with the acidic hydrogen of the N-H as the hydrogen donor. The spatial arrangement of molecules in the crystalline drug do not allow hydrogen bonding between the N-H and S=O to take place and thus the N-H bands are sharper and of higher intensity<sup>21</sup>.

#### **Dissolution studies**

A high amount of hydrophilic polymer may increase the availability of moisture, which may aid in devitrification<sup>26,27</sup>. The low-melting-point excipients such as PEGs and polyglycolized glycerides have been used widely as excipients in SD<sup>28,29</sup>. These excipients have increased drug dissolution rates by improving the wettability of the drug particles, by significant reducing the drug particle size during the formation of the SD, or by the inherently higher dissolution rate of the soluble component of the SD introducing the less-soluble component as finely divided particles into the dissolution medium30-32.

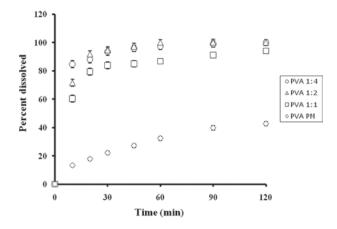


Figure 11. Dissolution profiles of the spray-dried particles of celecoxib with polyvinyl alcohol (PVA) 22000 and the physical mixture.

The dissolution profiles of the spray-dried particles and the PMs are presented in Figures 11-13. For SD containing PEG, complete dissolution in 900 mL occurred within 10 min for the 1:4 ratio and within 20 min for the 1:2 ratio (Figure 12). However, the SD with the 1:1 mass ratio required 45 min for complete dissolution. Only 59.94% of the celecoxib present in the PM was dissolved in 30 min. Table 3 presents the relative dissolution rate (RDR) of the different particles to that of pure celecoxib.

Since sufficient medium was present to allow sink conditions, dissolved drug did not reach saturation. The maximum amount of drug attained in the dissolution medium was 15.3 mg by the end of 2h. Complete dissolution of the celecoxib in the different PMs was never achieved in the dissolution study time. The lowest dissolution rate for celecoxib from a PM was observed in the case of PVA (Figure 11). Comparing this result with the dissolution rates of the spray-dried particles with 1:2 and 1:4 mass ratios with PVA, one can see the outstanding improvement in the dissolution rate of celecoxib following spray-drying.

Table 4 presents the efficiency with which the spraydrying process entraps celecoxib in the particles as well

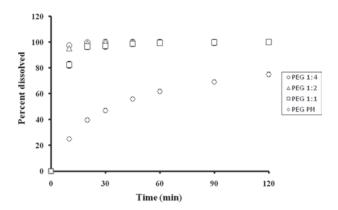


Figure 12. Dissolution profiles of the spray-dried particles of celecoxib with polyethylene glycol (PEG) 6000 and the physical mixture.

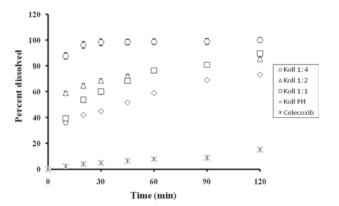


Figure 13. Dissolution profiles of the spray-dried particles of celecoxib with Kollicoat IR®, the physical mixture, and pure celecoxib.

Table 3. Relative dissolution rate (RDR) of celecoxib-polymer particles to that of pure celecoxib.

Polyvinyl alcohol (P	VA) 22000			
Time (min)	SD 1:1	SD 1:2	SD 1:4	Physical mixture
10	27.3	40.7	34.0	6.0
20	19.6	23.0	23.8	4.4
30	17.0	20.1	20.1	4.5
Polyethylene glycol	(PEG) 6000			
10	37.7	44.4	45.4	11.3
20	24.1	25.2	_	9.8
30	19.9	_	_	9.5
Kollicoat IR®				
10	17.9	26.8	42.9	15.9
20	13.3	16.0	25.7	10.4
30	12.2	13.9	_	9.0

All particles were passed through 200 µm sieve before dissolution.

Table 4. Spray-dried particulate properties.

Spray-dried		
particles	Production yield (%)	The actual drug load (%)
Polyvinyl alcol	nol (PVA) 22000	
SD 1:1	75.0	77.0
SD 1:2	75.0	71.6
SD 1:4	70.0	81.9
Polyethylene g	lycol (PEG) 6000	
SD 1:1	66.5	68.7
SD 1:2	75.0	79.3
SD 1:4	75.0	68.6
Kollicoat IR®		
SD 1:1	65.0	75.0
SD 1:2	78.0	76.0
SD 1:4	70.0	81.0

as the yield of the process. On the average, a 73% yield with an average of 76% entrapped drug can be noted. The lost amounts are attributed to the polymer allowing adherence to the wall of the spray-dryer and to the formation of very small particles that escaped with nitrogen gas and were lost to the filter. The 70% yield with 81% entrapment for celecoxib in a 1:4 mass ratio with either PVA or Kollicoat is a promising result.

An increased dissolution rate for celecoxib and a change from its crystalline form to the amorphous form can result from interactions with these polymers<sup>33,34</sup>. The spray-dried particles showed a spherical shape with both Kollicoat and PVA and a nearly spherical shape with a porous, spongy surface in the case of PEG, likely due to the presence of silicon dioxide. The small particle size provides a much greater overall surface area for dissolution and the highly hydrophilic polymers provide ready dissolution of the excipient<sup>35</sup>.

# **Conclusion**

Spray-drying is a good technique to utilize a hydrophilic polymer as a carrier in the formation of a SD. The promising results recommend the improvement of the dissolution rate of poorly water-soluble drugs using Kollicoat IR® or its constituents PVA or PEG. With the enhanced dissolution rate of celecoxib, categorized as class II in the Biopharmaceutical Classification System, its oral bioavailability is expected to increase. The potential for spray-dried particles with a spherical shape as well as the presence of an interaction between celecoxib and the polymer involved are important factors in the improvement of celecoxib dissolution rate.

# **Acknowledgements**

The authors gratefully acknowledge the Center of Excellence in Biotechnology Research (CEBR) for providing instruments and facilities to carry out this project.

#### Declaration of interest

This research was financially supported by the Research Centre at the College of Pharmacy, King Saud University, Riyadh, Saudi Arabia (grant number C.P.R.C. 251). The authors report no conflicts of interest. The authors alone are responsible for the content and writing of this article.

#### References

- Martin P, Bustamante A, Chun AHC. (1993). Physical Pharmacy. Philadelphia, PA: Lippincott Williams & Wilkins.
- 2. Geis GS. (1999). Update on clinical developments with celecoxib, a new specific COX-2 inhibitor: what can we expect? Scand J Rheumatol Suppl, 109:31-37.
- 3. Fort J. (1999). Celecoxib, a COX-2-specific inhibitor: the clinical data, Am I Orthop, 28:13-18.
- 4. Davies NM, Gudde TW, de Leeuw MA. (2001). Celecoxib: a new option in the treatment of arthropathies and familial adenomatous polyposis. Expert Opin Pharmacother, 2:139-152.
- Moore PA, Hersh EV. (2001). Celecoxib and rofecoxib. The role of COX-2 inhibitors in dental practice. J Am Dent Assoc, 132:451-456.
- 6. Tindall E. (1999). Celecoxib for the treatment of pain and inflammation: the preclinical and clinical results. J Am Osteopath Assoc, 99:S13-S17.
- 7. Tibble JA, Sigthorsson G, Foster R, Bjarnason I. (2000). Comparison of the intestinal toxicity of celecoxib, a selective COX-2 inhibitor,



- and indomethacin in the experimental rat. Scand J Gastroenterol, 35:802-807.
- 8. Ayalasomayajula SP, Kompella UB. (2004). Retinal delivery of celecoxib is several-fold higher following subconjunctival administration compared to systemic administration. Pharm Res, 21:1797-1804.
- 9. Paulson SK, Vaughn MB, Jessen SM, Lawal Y, Gresk CJ, Yan B et al. (2001). Pharmacokinetics of celecoxib after oral administration in dogs and humans: effect of food and site of absorption. J Pharmacol Exp Ther. 297:638-645.
- 10. Amrite AC, Ayalasomayajula SP, Cheruvu NP, Kompella UB. (2006). Single periocular injection of celecoxib-PLGA microparticles inhibits diabetes-induced elevations in retinal PGE2, VEGF, and vascular leakage. Invest Ophthalmol Vis Sci, 47:1149-1160.
- 11. Amidon GL, Lennernäs H, Shah VP, Crison JR. (1995). A theoretical basis for a biopharmaceutic drug classification: the correlation of in vitro drug product dissolution and in vivo bioavailability. Pharm Res. 12:413-420
- 12. Banga S, Chawla G, Varandani D, Mehta BR, Bansal AK. (2007). Modification of the crystal habit of celecoxib for improved processability. J Pharm Pharmacol, 59:29-39.
- 13. Subramanian N, Ray S, Ghosal SK, Bhadra R, Moulik SP. (2004). Formulation design of self-microemulsifying drug delivery systems for improved oral bioavailability of celecoxib, Biol Pharm Bull, 27:1993-1999.
- 14. Dixit RP, Nagarsenker MS. (2007). In vitro and in vivo advantage of celecoxib surface solid dispersion and dosage form development. Indian J Pharm Sci, 69:370-377.
- 15. Nagarsenker MS, Joshi MS. (2005). Celecoxib-cyclodextrin systems: characterization and evaluation of in vitro and in vivo advantage. Drug Dev Ind Pharm, 31:169-178.
- 16. Gupta VR, Mutalik S, Patel MM, Jani GK. (2007). Spherical crystals of celecoxib to improve solubility, dissolution rate and micromeritic properties. Acta Pharm, 57:173-184.
- 17. Guzmán HR, Tawa M, Zhang Z, Ratanabanangkoon P, Shaw P, Gardner CR et al. (2007). Combined use of crystalline salt forms and precipitation inhibitors to improve oral absorption of celecoxib from solid oral formulations. J Pharm Sci, 96:2686-2702.
- 18. Pouton CW. (2006). Formulation of poorly water-soluble drugs for oral administration: physicochemical and physiological issues and the lipid formulation classification system. Eur J Pharm Sci, 29:278-287.
- 19. Kolter K. (2002). Kollicoat IR®—innovation in instant release film coating. BASF ExAct, 8:4-5.
- 20. Punitha S, Karthikeyan D, Devi P, Vedha Hari BN. (2009) Enhancement of solubility and dissolution of celecoxib by solid dispersion technique. J Pharm Sci Technol, 2:63-68.

- 21. Chawla G, Gupta P, Thilagavathi R, Chakraborti AK, Bansal AK. (2003). Characterization of solid-state forms of celecoxib. Eur J Pharm Sci, 20:305-317.
- 22. Thimmasetty J, Subrahmanyam CVS, Vishwanath BA, Sathesh Babu PR. (2009). Solubility parameter estimation of celecoxib by current methods. Asian J Res Chem, 2:188-195.
- 23. Vasu Dev R, Shashi Rekha K, Vyas K, Mohanti SB, Rajender Kumar P, Om Reddy G. (1999). Celecoxib, a COX-II inhibitor. Acta Cryst, C55:IUC9900161 [doi:10.1107/S0108270199098200]
- 24. Qian XF, Yin J, Guo XX, Yang YF, Zhu ZK, Lu J. (2000). Polymerinorganic nanocomposites prepared by hydrothermal method: PVA/ZnS, PVA/CdS, preparation and characterization. J Mater Sci Letters, 19:2235-2237.
- 25. Biswal S, Sahoo J, Murthy PN. (2009). Characterization of gliclazide-PEG 8000 solid dispersions. Trop J Pharm Res, 8:417-424.
- 26. Gupta P, Bansal AK. (2005). Devitrification of amorphous celecoxib. AAPS Pharmscitech, 6:E223-E230.
- 27. Puria V, Dantuluria AK, Kumarb M, Kararb N, Bansal AK. (2010). Wettability and surface chemistry of crystalline and amorphous forms of a poorly water soluble drug. Eur J Pharm Sci, 40:84-93.
- 28. Yassin AE, Alanazi FK, El-Badry M, Alsarra IA, Barakat NS, Alanazi FK. (2009). Preparation and characterization of spironolactoneloaded gelucire microparticles using spray-drying technique. Drug Dev Ind Pharm, 35:297-304.
- 29. Shimpi SL, Mahadik KR, Paradkar AR. (2009). Study on mechanism for amorphous drug stabilization using gelucire 50/13. Chem Pharm Bull, 57:937-942.
- 30. Dordunoo SK, Ford JL, Rubinstein MH. (1991). Preformulation studies on solid dispersions containing triamterene or temazepam in polyethylene glycols or gelucire 44/14 for liquid filling of hard gelatin capsules. Drug Dev Ind Pharm, 17:1685-1713.
- 31. Leuner C, Dressman J. (2000). Improving drug solubility for oral delivery using solid dispersions. Eur J Pharm Biopharm, 50:47-60.
- 32. Passerini N, Perissutti B, Moneghini M, Voinovich D, Albertini B, Cavallari C et al. (2002). Characterization of carbamazepinegelucire 50/13 microparticles prepared by a spray-congealing process using ultrasounds. J Pharm Sci, 91:699-707.
- 33. Hancock BC, Zografi G. (1997). Characteristics and significance of the amorphous state in pharmaceutical systems. J Pharm Sci, 86:1-12.
- 34. El-Badry M, Fetih G, Fathy M. (2009). Improvement of solubility and dissolution rate of indomethacin by solid dispersions in gelucire 50/13 and PEG4000. Saudi Pharm J, 17:217-225
- 35. Karavas E, Georgarakis E, Sigalas MP, Avgoustakis K, Bikiaris D. (2007). Investigation of the release mechanism of a sparingly water-soluble drug from solid dispersions in hydrophilic carriers based on physical state of drug, particle size distribution and drug-polymer interactions. Eur J Pharm Biopharm, 66:334-347.